Haemostatic Profile in Patients of Myeloproliferative Neoplasms-A Tertiary Care Centre Experience

YATENDRA PARASHAR¹, RASHMI KUSHWAHA², ASHUTOSH KUMAR³, KAMAL AGARWAL⁴, U.S SINGH⁵, MILI JAIN⁶, S.P VERMA⁷, A.K TRIPATHI⁸

ABSTRACT

Pathology Section

Introduction: Patients of Myeloproliferative Neoplasm (MPN) commonly present with abnormalities in laboratory coagulation tests that are consistent with hypercoagulable state. Some individuals with MPN exhibit a pattern of exclusive bleeding or thrombotic events; many others have both bleeding and thrombosis during the course of the disease.

Aim: This study was undertaken to assess the haemostatic defects and platelet functions in patients of Myeloproliferative neoplasms.

Materials and Methods: One year prospective study was conducted at a tertiary care centre in North India in Department of Pathology in collaboration with Department of Clinical Haematology. All recently diagnosed cases of MPN along with 30 age and sex matched controls were included. Patients on antiplatelet drugs, antimyeloproliferative treatment, vitamin K agonists or antagonists, OCP's, Platelet count <1,00, 000/µl, high grade fever, liver disease, pregnancy were excluded from this study.

All the patients underwent screening investigations like CBC, peripheral smear evaluation, bleeding time, prothrombin

time, Activated Partial Thromboplastin Time, Protein C and S measurement (clot based assay) and aggregation studies with ADP, 5μ M (Optical Aggregometry with AGGRO/LINK 8 software and CHRONOLOG 700 aggregometer).

Results: In present study, 50 cases were included. There was an occult prothrombotic state, suggested by significantly (p<0.001) reduced levels of Protein C and Protein S, but no patient presented with frank thrombosis while 8 out of 50 patients had haemorrhagic manifestations ranging from subdural haematoma to pin point petechial haemorrhages. Patients of CML-CP, ET, PV, PMF, MPN-NOS showed significantly reduced maximal aggregation with 5µM ADP when compared to control (p<0.001). MPV also showed a statistically significant increase in these patients.

Conclusion: Thrombohaemorrhagic complications significantly affect the morbidity and mortality of MPN patients. This can be assessed by the use of platelet aggregation studies, protein C and S activities and other coagulation studies. Timely diagnosis of these prothrombotic/haemorrhagic states can decrease the morbidity in these patients.

Keywords: Haemorrhage, Platelet dysfunction, Thrombosis

INTRODUCTION

Life expectancy of patients with Myeloproliferative neoplasms is strongly compromised by the haemostatic complications, like vascular thrombosis and haemorrhages [1]. These patients present with arterial or venous thrombosis. Arterial thrombosis accounts for 60%-70% of events related to myeloproliferative neoplasms and it includes ischemic stroke, acute myocardial infarction, and peripheral arterial occlusion [2].

These patients can also present with deep venous thrombosis of the lower extremities, pulmonary embolism, intra abdominal (hepatic, portal and mesenteric) and cerebral vein thrombosis. In polycythemia vera (PV) patients, venous thrombosis is relatively common and constitutes approximately one-third of total events. Many a times splanchnic and cerebral vein thrombosis is the presenting feature in patients of MPN. Also, the prevalence of splanchnic and cerebral vein thrombosis is higher in these patients [2]. Thrombotic complications not only involve large blood vessels but also involve microvasculature causing digital ischemia, pregangrenous changes, erythromelalgia, dizziness, confusion, headache, seizures and visual disturbances [3].

Patients of MPN are in a hypercoagulable state even in the absence of frank thrombotic manifestations. This is proven by the increased levels of plasma biomarkers of haemostatic system activation [4-6].

This study was undertaken to assess the haemostatic defects and platelet functions in patients of Myeloproliferative neoplasms.

MATERIALS AND METHODS

This was a prospective study of one year duration from (August 2014 to July 2015) conducted in a tertiary care centre in North India. Fifty patients of Myeloproliferative neoplasm diagnosed after peripheral smear and bone marrow evaluation were included in this study. Patients taking vitamin K, antiplatelet/antithrombotic drugs, antimyeloproliferative drugs, patients with liver disease, high fever, patients on OCP's and pregnant patients were excluded from this study. Apart from this, 30 age and sex matched normal healthy individuals who were not receiving drugs interfering with platelet functions or coagulation were selected as controls.

All the patients underwent screening investigations like CBC, peripheral smear evaluation, Bleeding Time (BT), Prothrombin Time (PT), Activated Partial Thromboplastin Time (aPTT), Protein C and S measurement (clot based assay using reagents from Tulip Diagnostics India) and aggregation studies with ADP 5µM (Optical Aggregometry with AGGRO/LINK 8 software and CHRONOLOG 700 aggregometer).

Continuous data were summarized as Mean \pm SD (standard deviation) while discrete (categorical) in number and percentage (%). Continuous groups were compared by Student's t-test. Categorical groups were compared by chi-square (χ^2) test. A two-tailed p<0.05 was considered statistically significant. Analyses were performed on SPSS software (Windows version 17.0). Ethical clearance was obtained for this study.

RESULTS

A total of 50 cases were included in present study, out of which 42 were of Chronic Myeloid Leukaemia (CML), 2 each of Chronic Neutrophilic Leukaemia (CNL), Esential Thrombocythemia (ET), Primary Myelofibrosis (PMF) and one each of Polycythemia Vera (PV) and MPN-nos. [Table/Fig-1] is showing the age distribution of cases. In the present study, out of 50 patients, 16 (32.0%) were females and 34 (68%) were males. Both patients of ET and single patient of PV were males. 69% patients of CML were males, while sex distribution in CNL and PMF was 1:1. Thirty Age and sex matched control were included.

Out of 50 patients, only 8 patients had haemorrhagic manifestations. None of patients presented with clinical frank thrombosis.

Haemorrhagic manifestations noted were epistaxis, melena, menorrhagia, gum bleeding, haematoma and petechial rashes. One patient of CML presented with subdural haematoma and one ET patient presented with petechial rashes mostly on lower limbs and forearms.

Difference in values for all the coagulation variables including platelet aggregation, in control and cases was found to be statistically significant. Values of Maximal Aggregation with ADP 5μ M, Protein C and Protein S in cases were found to be lower than that of Controls while values of BT, PT, aPTT, MPV and PDW was found to be higher in cases as compared to controls [Table/ Fig-2].

Maximal platelet aggregation, Protein C and S were found to be significantly lower in our patients of MPN as compared to controls (p<0.001). Mean platelet volume (MPV) was found to be significantly higher in our MPN cases as compare to controls (p<0.001).

Values of Platelet Distribution Width (PDW) showed that patients of Myeloproliferative disorders have higher variation in size of their platelets. This variation was found to be more in Essential

Age Group (years)	CML- (CP+BP)	CNL	ET	MPN- NOS	PMF	PV
Upto 10	2	-	-	-	-	-
11-20	1	-	-	-	-	-
21-30	6	-	-	-	-	-
31-40	20	-	1	1	-	-
41-50	10	1	1	-	1	-
>50	3	1	-	-	1	1
Total	42	2	2	1	2	1

[Table/Fig-1]: Number of patents in each age group.

Variables	Control (n=30)	Cases (n=50)	Statistical significance (Student t-test)				
	Mean±SD.	Mean±SD	't'	ʻp'			
Bleeding time (BT) (in minutes)	2.73±0.64	4.74±2.90	-3.724	<0.001			
Activated partial thromboplastin time (aPTT) (in seconds)	28.30±5.59	32.00±5.23	-2.984	0.004			
Prothrombin time (PT) (in seconds)	12.50±1.36	13.41±1.67	-2.522	0.014			
Maximal aggregation (%)	78.97±5.31	52.57±23.40	6.072	<0.001			
Protein C (% activity)	96.03±10.20	60.67±22.88	7.987	<0.001			
Protein S (% activity)	104.2±12.72	58.40±25.88	9.044	<0.001			
Mean platelet volume MPV(fl)	8.73±1.10	10.45±1.67	-5.008	<0.001			
Platelet distribution width PDW (%)	17.23±2.40	29.48±20.31	-3.282	0.002			
[Table/Fig-2]: Comparison of various coagulation parameters between Controls							

-					
~					
_			-		
				7	7

Variables	No haemoi (n=4	n- rrhagic 42)	Haemo (n=	errhagic =8)	Statistical Significance	
	Mean	± SD	Mean	± SD	t	ʻp'
Bleeding Time (minute)	3.71	1.73	10.12	1.42	9.8420	<0.0001
aPTT (second)	31.57	5.06	34.25	5.90	1.3384	0.1871
PT (second)	13.39	1.69	13.50	1.69	0.1687	0.8667
Platelet count (cumm)	290381	142181	370000	253068	1.2653	0.2119
MPV(fl)	10.49	1.76	10.23	1.07	0.4019	0.6896
PDW(%)	27.93	18.25	37.60	29.09	1.2412	0.2206
Maximal aggregation (ADP 5µM (%age)	52.21	23.99	54.45	21.39	0.2458	0.8069
Protein C (% age activity)	61.70	23.24	55.25	21.43	0.7274	0.4705
Protein S (%age activity)	60.06	25.63	49.66	27.15	1.0426	0.3023

[Table/Fig-3]: Comparison between Non-haemorrhagic and Haemorrhagic Cases.



[Table/Fig-4]: CT scan - chronic subdural hemorrhage in fronto-parietal region.

		Maximal percentage aggregation (%)						
MPN-subtype	of cases	Mean Std. dev. (± SD)		Minimum	Maximum			
CML-BP	1	79.00	-	79	79			
CML-CP	41	55.02	22.81	12	105			
CNL	2	67.00	2.83	65	69			
ET	2	26.30	13.43	17	36			
PV	1	44.00	-	44	44			
PMF	2	20.00	4.24	17	23			
MPN-NOS	1	23.00	-	23	23			
Table / Eig 51: Subtypes of MDN and Maximal District Aggregation (5UM ADD)								

Thrombocythemia patients; such variations represented as high

values of standard deviation in our patients of MPN. Further differences in mean PDW of cases and controls were highly significant with p-value <0.01.

Out of 50 cases, only 8 patients had haemorrhagic manifestations. Difference between bleeding time of haemorrhagic and non haemorrhagic cases was statistically significant (p<0.0001) [Table/ Fig-3]. Haemorrhagic manifestations seen in seven patients of CML included episodes of epistaxis, melena, menorrhagia, gum bleeding haematoma and petechial rashes. One patient of CML presented with chronic subdural haemorrhage [Table/Fig-4].

Other variables showed only small differences between haemorrhagic and non-haemorrhagic MPN patients. Patients of CML-CP, ET, PV, PMF, MPN-NOS had reduced maximal aggregation with 5μ M ADP [Table/Fig-5,6].

DISCUSSION

Thrombohaemorrhagic complications in MPN are common morbid conditions. In present study there was an occult prothrombotic



state, but no patient presented with frank thrombosis while eight out of fifty patients had haemorrhagic manifestations ranging from subdural haematoma to pin point petechial haemorrhages.

In present study, the only patient of PV, presented with symptoms suggestive of microcirculatory disturbances (headache, dizziness, pruritus) but no frank thrombosis was seen.

One of ET patients presented with petechial rashes on the lower limbs; suggestive of qualitative platelet defect or defective vascular endothelial cells. Patients of PMF, CNL and MPN–NOS didn't show any thrombotic or haemorrhagic manifestations.

Another inference which can be made is that patients with normal or even elevated platelet count can present with raised BT and haemorrhagic manifestations; this finding gives us an insight of defective platelet functions in MPN as stated by previous workers [7-14]. Pathogenesis behind such findings includes increased expression of P-selectin and tissue factors on platelet in MPN patients which may lead to exhaustion of functional ability of platelets to aggregate. Another paradox is that very high number of platelets as seen in ET patients (8,73,000 and 5,60,000/mm³) may consume all available large circulatory vWF multimers and results in increased bleeding time and haemorrhagic manifestations [15].

In present study, there were significantly reduced levels of maximal platelet aggregation with ADP (5.0μ M) ($52.57\pm23.40\%$) as compared to healthy controls ($78.97\pm5.31\%$) with p<0.001(see [Table/Fig-2]). Such results have been found in many studies as mentioned in [Table/Fig-7] [16-20]. All of these studies emphasize on the loss of secondary wave of aggregation and reduced maximal platelet aggregation.

These abnormalities can be justified by the defective arachidonic acid metabolism, reduced activation of platelets by agonists, acquired storage pool disease, decreased number of α^2 -adrenergic receptors, altered expression of specific platelet membrane glycoproteins (GP) such as GPIIb/IIIa, GP Ib/IX or increased number of GPIV molecules and increased expression of receptors for the Fc component of IgG, acquired von Willebrand disease, a reduction of platelet procoagulant activity and an acquired form of Bernard-Soulier syndrome as reported in these disorders by other researchers [7-23].

In present study, activity of Protein C was found to be reduced very significantly [Tables/Fig-2,3] in MPN Cases (60.67±22.88%) as compare to the mean activity in Controls (96.03±10.20%) with p<0.001. Similar findings are seen in studies done by other workers and they found an acquired activated protein C resistance in MPN Cases [Table/Fig-8] [24-26].

Low levels of Protein C indicate the presence of "Prothrombotic State" in MPN patients even in the absence of frank thrombosis.

Protein S activity was also found to be reduced in MPN cases (see [Table/Fig-2]) (58.40±25.88%) in the present study as compare to activity of protein S in healthy controls (104.20±12.72%).

Such kind of differences have also been seen by other researchers. These results can be justified by the researcher's that indicate that proteases from platelets and leukocytes are able to cleave PS in plasma [27]. Separate studies done by Marchetti M et al., and Falanga et al., showed that increased neutrophil elastase levels in MPN patients degrade the protein S. Studies also showed that factor V also undergo similar degradation by elastases [28-30].

Falanga et al., suggested that increased blood coagulation in MPN patients is multifactorial and it involves abnormalities in platelets, erythrocytes, leucocytes and endothelial cells. Sekhar M et al., found splenic vein thrombosis to be one of the most common presentations of MPN. However, in our studies no such cases were found. Martinelli I et al., found that MPN patients are at increased risk of cerebral vein thrombosis [31-33].

Andhan	Platel	Patient						
Author	ADP		Collagen		Epinephrine		population	
Jesus M Cesar (2005) [16]	6/55	11%	21/55	38%	32/55	58%	ET only	
SimonaAvram (2001) [17]	19/76	25%	-	-	-	-	All MPN	
Waddell CC (1981) [18]	9/18	50%	7/18	38.9%	11/18	61.1%	All MPN	
Pareti FI (1982) [19]	19/52	36.5%	9/52	17.3%	15/52	28.8%	All MPN	
Russell NH (1981) [20]	9/17	52.9%	10/17	58.8%	-	-	All MPN	
Present study	40/50	80%	-	-	-	-	All MPN	
[Table/Fig-7]: Comparison of platelet aggregation (%) in different studies and in present study [16-20]								

Author	Number of patients with deficiencies of PC and PS	Percentage					
Bucalossi (1996) [24]	22/81	27.16%					
L Amitrano (2003) [25]							
Protein C	7/61	11.4%					
Protein S	7/79	8.8%					
Hoekstra J(2011) [26]	4/14	28.6%					
Present study							
Protein C	34/50	68%					
Protein S	30/50	60%					
[Table/Fig-8]: Comparison of Protein C & S activity in different studies [24-26].							

MPV of MPN cases (10.45 \pm 1.67fl) was found to be statistically higher in comparison with that of controls (8.73 \pm 1.10) having p <0.001 [Table/Fig-2]. These higher values of MPV are suggestive of presence of large sized platelets in MPN patients.

PDW of MPN Cases (29.48 \pm 20.31%) was compared with that of controls (17.23 \pm 2.40%) and was found to be significantly higher in MPN cases (p<0.01) [Table/Fig-2].

Higher value of PDW in MPN cases is an indicator of presence of variable sized platelets in MPN patients as seen in various studies [34,35].

In present study, MPV was correlated with Maximal Platelet Aggregation (categorized as <70% and \geq 70%); a statistically significant correlation was found (p<0.05) as patients with higher MPV had reduced Maximal Platelet Aggregation.

LIMITATIONS

This study was of short duration and effect of MPN treatment on coagulation profile was not studied.

CONCLUSION

Thrombohaemorrhagic complications significantly affect the morbidity and mortality of MPN patients. These are multifactorial events that include the key role of platelets with their in vivo activation and role of vascular endothelial cells, leukocytes, and reduced activity of protein C and S. Also, these patients are in a continuous hypercoagulable state even in the absence of frank thrombosis. Hence, patients of MPN, should be monitored and councelled appropriately keeping these complications in view.

REFERENCES

- Tefferi A. Polycythemia vera and essential thrombocythemia: 2012 update on [1] diagnosis, risk stratification, and management. Am J Haematol. 2012;87(3):285-93.
- [2] Reikvam H, Tiu RV. Venous thromboembolism in patients with essential thrombocythemia and polycythemia vera. Leukaemia. 2012;26(4):563-71.
- Pearson TC, Humphrey PRD, Thomas DJ, Wetherley-Mein G. Haematocrit, [3] blood viscosity, cerebral blood flow, and vascular occlusion. Springer - Verlag. 1981;97-107
- Falanga A, Marchetti M, Evangelista V, Vignoli A, Licini M, Balicco M, et al. [4] Polymorphonuclear leukocyte activation and haemostasis in patients with essential thrombocythemia and polycythemia vera. Blood. 2000;96(13):4261-66.
- Arellano-Rodrigo E, Alberto Alvarez-Larraín, Reverter JC, Colomer D, Villamor [5] N, Bellosillo B, et al. Platelet turnover, coagulation factors, and soluble markers of platelet and endothelial activation in essential thrombocythemia: relationship with thrombosis occurrence and JAK2 V617F allele burden. Am J Haematol. 2009;84(2):102-08.
- [6] Landolfi R, Rocca B, Patrono C. Bleeding and thrombosis in myeloproliferative disorders: mechanisms and treatment. Crit Rev Oncol Haematol. 1995;20(3):203-22
- [7] Wehmeier A, Tschope D, Esser J, Menzel C, Nieuwenhuis HK, SchneiderW. Circulating activated platelets in myeloproliferative disorders. Thromb Res. 1991;61(3):271-78.
- Ireland H, Lane DA, Wolff S, Foadi M. In vivo platelet release in myeloproliferative [8] disorders. Thromb. Haemost. 1982;48(1):41-45.
- [9] Mazzucato M, De Marco L, De Angelis V, De Roia D, Bizzaro N, Casonato A. Br J Haematol, 1989:73(3):369-74.
- Clezardin P, McGregor JL, Dechavanne M, Clemetson KJ. Platelet membrane [10] glycoprotein abnormalities in patients with myeloproliferative disorders and secondary thrombocytosis. Br J Haematol. 1985;60(2):331-44.
- [11] Lopez Fernandez MF, Lopez Berges C, Martin R, Abnormal structure of von Willebrand factor in myeloproliferative syndrome in associated to either thrombotic or bleeding diathesis. Thromb Haemost.1987;58(2):753-57.
- Budde U, Dent JA, Berkowitz SD. Subunit composition of plasma von Willebrand [12] factor in patient with the myeloproliferative syndrome. Blood. 1986;68(6):1213-17.
- [13] Berndt MC, Kabral A, Grimsley P, An acquired Bernard-Soulier-like platelet defect associated with juvenile myeloproliferative syndrome. Br J Haematol. 1988;68:97-101.
- [14] Hehlmann R, Jahn M, Baumann, Kopcke W. Essential thrombocythemia. Clinical characteristics and course of 61 cases. Cancer. 1988;61(12):2487-96.
- Landolfi R, Cipriani MC, Novarese L. Thrombosis and bleeding in polycythemia [15] vera and essential thrombocythemia: pathogenetic mechanisms and prevention. Best Pract Res Clin Haematol. 2006;19(3):617-33.
- [16] Cesar JM, de Miguel D, García Avello A, Burgaleta C. Platelet Dysfunction in Primary Thrombocythemia Using the Platelet Function Analyzer, PFA-100. Am J Clin Pathol. 2005;123:772-77.

- [17] Avram S, Lupu A, Angelescu S, Olteanu N, Mut-Popescu D. Abnormalities of platelet aggregation in chronic myeloproliferative disorders. J Cell Mol Med. 2001; 5(1):79-87.
- [18] Waddell CC, Brown JA, Repinecz YA. Abnormal platelet function in myeloproliferative disorders. Arch Pathol Lab Med. 1981;105(8):432-35
- [19] Pareti Fl, Gugliotta L, Mannucci L, Guarini A, Mannucci PM. Biochemical and metabolic aspects of platelet dysfunction in chronic myeloproliferative disorders. Thromb Haemostas. 1982;47:84-89.
- [20] Russell NH, Salmon J, Keenan JP, Bellingham AJ. Platelet adenine nucleotides and arachidonic acid metabolism in the myeloproliferative disorders. Thromb Res. 1981;22(4):389-97.
- [21] Mayordomo O, Carcamo C, Vecino AM. Arachidonic acid metabolism in platelets of patients with essential thrombocythemia. Thromb Res. 1995;78(4):315-21.
- [22] Takayama H, Okuma M, Kanaji K. Altered arachidonate metabolism by leukocytes and platelets in myeloproliferative disorders. Prostaglandins Leukot Med. 1983;12(3):261-72.
- Smith I.L, Martin TJ. Platelet thromboxane synthesis and release reactions in [23] myeloproliferative disorders. Haemostasis. 1982;11(2):119-27.
- [24] Bucalossi A, Marotta G, Bigazzi C, Galieni P, Dispensa E. Reduction of antithrombin III, protein C, and protein S levels and activated protein C resistance in polycythemia vera and essential thrombocythemia patients with thrombosis. Am J Haematol. 1996;52(1):14-20.
- Amitrano L, Guardascione, Ames PRJ, Margaglione M, Antinolfi I, Iannaccone [25] L, et al. Thrombophilic Genotypes, Natural Anticoagulants, and Plasma Homocysteine in Myeloproliferative Disorders: Relationship With Splanchnic Vein Thrombosis and Arterial Disease. Am J Haematol. 2003;72:75-81.
- Hoekstra J, Bresser EL, Smalberg JH, Spaander MC, Leebeek FW, Janssen HL. [26] Long-term follow-up of patients with portal vein thrombosis and myeloproliferative neoplasms. J Thromb Haemost. 2011;9(11):2208-14.
- [27] Brinkman HJ, Mertens K, van Mourik JA. Proteolytic cleavage of protein S during the haemostatic response. J Thromb Haemost. 2005;3(12):2712-20.
- [28] Marchetti M, Castoldi E, Spronk HM, van Oerle R, Balducci D, Barbui T, et al. Thrombin generation and activated protein C resistance in patients with essential thrombocythemia and polycythemia vera. Blood. 2008;112:4061-68
- [29] Eckle I, Seitz R, Egbring R, Kolb G, Havemann K. Protein S degradation in vitro by neutrophil elastase. Scand J Clin Lab Invest. 1993;53(3):281-88.
- [30] Falanga A, Marchetti M, Vignoli A, Balducci D, Russo L, Guerini V, et al. V617F JAK-2 mutation in patients with essential thrombocythemia: relation to platelet, granulocyte, and plasma haemostatic and inflammatory molecules. Experimental haematology. 2007;35:702-11.
- [31] Falanga A, Marchetti M. Thrombosis in myeloproliferative neoplasms. Semin Thromb Haemost. 2014;40(3):348-58.
- [32] Sekhar M, McVinnie K, Burroughs ak. Splanchnic vein thrombosis in myeloproliferative neoplasms. British Journal of Haematology. 2013;162:730-47.
- [33] Martinelli I, De Stefano V, Carobbio A, Randi ML, Santarossa C, Rambaldi A, et al. Cerebral vein thrombosis in patients with Philadelphia-negative myeloproliferative neoplasms. An European Leukaemia Net study. Am J Haematol. 2014;89(11):E200-05.
- [34] Murphy S. Thrombocytosis and thrombocythemia. Clin Haematol. 1983;12:89-102.
- Zeigler Z, Murphy S, Gardner FH. Microscopic platelet size and morphology in [35]
- various haematologic disorders. Blood. 1978;51(3):479-86

PARTICULARS OF CONTRIBUTORS:

- Junior Resident, Department of Pathology, King George's Medical University, Lucknow, Uttar Pradesh, India.
- 2. Associate Professor, Department of Pathology, King George's Medical University, Lucknow, Uttar Pradesh, India.
- Professor and Head, Department of Pathology, King George's Medical University, Lucknow, Uttar Pradesh, India. 3
- 4 Professor, Department of Pathology, King George's Medical University, Lucknow, Uttar Pradesh, India.
- Professor, Department of Pathology, King George's Medical University, Lucknow, Uttar Pradesh, India. Assistant Professor, Department of Pathology, King George's Medical University, Lucknow, Uttar Pradesh, India. 5
- 6.
- Assistant Professor, Department of Clinical Hematology, King George's Medical University, Lucknow, Uttar Pradesh, India.
- 8 Head of Department of Clinical Hematology, King George's Medical University, Lucknow, Uttar Pradesh, India.

NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

Dr. Rashmi Kushwaha,

Associate Professor, Lymphoma-Leukemia Lab, Department of Pathology, K.G.M.U, Lucknow-226003, Uttar Pradesh, India. E-mail: docrashmi27@yahoo.co.in

Date of Submission: Mar 08, 2016 Date of Peer Review: Apr 28, 2016 Date of Acceptance: Aug 10, 2016 Date of Publishing: Nov 01, 2016

FINANCIAL OR OTHER COMPETING INTERESTS: None.